

# Stomatal response of *Pinus sylvestris* to elevated CO<sub>2</sub> concentrations during the four years of exposure

ZHOU Yu-mei, HAN Shi-jie\*, LIU Ying, JIA Xia

*Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang 110016, P. R. China*

**Abstract:** Four-year-old *Pinus sylvestris* were exposed for four growing seasons in open top chambers to ambient CO<sub>2</sub> concentration (approx. 350  $\mu\text{mol} \cdot \text{mol}^{-1}$ ) and high CO<sub>2</sub> concentrations (500 and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$ ) at Research Station of Changbai Mountain Forest Ecosystems, Chinese Academy of Sciences at Antu Town, Jilin Province, China (42°N, 128°E). Stomatal response to elevated CO<sub>2</sub> concentrations was examined by stomatal conductance ( $g_s$ ), ratio of intercellular to ambient CO<sub>2</sub> concentration ( $c_i/c_a$ ) and stomatal number. Reciprocal transfer experiments of stomatal conductance showed that stomatal conductance in high-[CO<sub>2</sub>]-grown plants increased in comparison with ambient-[CO<sub>2</sub>]-grown plants when measured at their respective growth CO<sub>2</sub> concentration and at the same measurement CO<sub>2</sub> concentration (except a reduction in 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> grown plants compared with plants on unchambered field when measured at growth CO<sub>2</sub> concentration and 350  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>). High-[CO<sub>2</sub>]-grown plants exhibited lower  $c_i/c_a$  ratios than ambient-[CO<sub>2</sub>]-grown plants when measured at their respective growth CO<sub>2</sub> concentration. However,  $c_i/c_a$  ratios increased for plants grown in high CO<sub>2</sub> concentrations compared with control plants when measured at the same CO<sub>2</sub> concentration. There was no significant difference in stomatal number per unit long needle between elevated and ambient CO<sub>2</sub>. However, elevated CO<sub>2</sub> concentrations reduced the total stomatal number of whole needle by the decline of stomatal line and changed the allocation pattern of stomata between upper and lower surface of needle.

**Key words:**  $c_i/c_a$  ratio; High CO<sub>2</sub>; *Pinus sylvestris*; Stomatal conductance; Stomatal number; Stomatal line

Abbreviations:  $g_s$ , stomatal conductance;  $c_i$ , intercellular CO<sub>2</sub> concentration;  $c_a$ , ambient CO<sub>2</sub> concentration

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## Introduction

Stomata directly affect the gas exchange of CO<sub>2</sub>/H<sub>2</sub>O between atmosphere and foliage. The impact of elevated CO<sub>2</sub> concentration on the stomatal behavior has attracted considerable attention. The stomatal response to CO<sub>2</sub> is important in understanding stomatal physiology, and vegetation-atmosphere exchanges at all scales from the individual plant up to global vegetation (Morison 1998). Short-term and long-term effects of increased CO<sub>2</sub> on stomata are different. In general, the short-term response of stomata is a change in aperture (usually reversible), and long-term response includes anatomical and morphological changes, for example, in stomatal number and/or in size (Morison 1998). Stomatal acclimation may occur when plants are exposed to increased CO<sub>2</sub> concentration for a long time.

Stomatal conductance ( $g_s$ ) is most frequently used for assessing the function of stomata in reconciling the water loss and carbon gain (Zhang *et al.* 2002). It is generally accepted that an increase in the ambient CO<sub>2</sub> concentration can cause reductions in stomatal conductance resulted from the decrease of stomatal aperture and/or density, and the reduction varied widely (Bunce 2000; Morison 2001). A reduction in stomatal conductance is a common response of herbaceous plants to elevated CO<sub>2</sub> (Bunce 2000). Some experimental evidences suggested that many forest tree species show small or non-significant change in stomatal

conductance under long-term elevated CO<sub>2</sub> (Curtis 1996; Saxe *et al.* 1998), particular conifer (Teskey 1995).

Stomata appear to response directly to the intercellular CO<sub>2</sub> concentration ( $c_i$ ), rather than ambient CO<sub>2</sub> concentration ( $c_a$ ), as demonstrated by Mott (Mott 1988). C<sub>3</sub> plants normally maintain relative constancy of the ratio of intercellular to ambient CO<sub>2</sub>, approx. 0.7 (Lodge *et al.* 2001). Given no adjustment of stomata, the rate of CO<sub>2</sub> diffusion through the stomatal pores would rise in proportion to the increase in ambient CO<sub>2</sub> (Jarvis *et al.* 1999). Therefore, whether  $c_i/c_a$  ratio remains constant with increased  $c_a$  should be examined carefully.

The change of stomatal number of needle is a long-term response to elevated CO<sub>2</sub> concentration. Some literatures have reported no change in stomatal density (Poole *et al.* 2000; Lodge *et al.* 2001). There was no difference in stomata density for current-year needles of Sitka spruce trees exposed to elevated CO<sub>2</sub> concentration for 4 years between treatments (Barton and Jarvis, 1999), the same phenomenon also was observed on *Alnus glutinosa* (Poole *et al.* 2000).

The main objectives of this study are to determine stomatal response of *Pinus sylvestris* to long-term exposure to high CO<sub>2</sub> concentration: (a) to determine  $g_s$  and  $c_i/c_a$  ratio at different measurement of CO<sub>2</sub> concentration; (b) to examine the changes of stomatal number of current-year needle.

## Materials and methods

The study site is located at Research Station of Changbai Mountain Forest Ecosystems, Chinese Academy of Sciences at Antu Town, Jilin Province, China (42°N, 128°E). Average annual rainfall is 700 mm. In 1999, seedlings of *Pinus sylvestris* were planted in open top chambers and on un-chambered field. Open top chamber consists of aluminium frames of 1.2 m in length, 0.9 m in width and height, and clear glass covers. CO<sub>2</sub> enters the chamber through perforated plastic pipe at the bottom

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**Biography:** ZHOU Yu-mei (1973- ), female, Ph. Doctor, assistant research fellow, Institute of Applied Ecology, Chinese Academy of Sciences, Shenyang 110016, P. R. China. E-mail: zhouyumei73@126.com

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\*Corresponding author

of chamber. Fan is hung in the top of the chamber to mix the gas well-proportioned. Treatments consist of three concentration levels of CO<sub>2</sub>: ambient, 500 and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>. Seedlings in the control chamber and on un-chambered field are given ambient CO<sub>2</sub>, approx. 350  $\mu\text{mol} \cdot \text{mol}^{-1}$ . The CO<sub>2</sub> concentrations in each chamber were checked weekly and adjusted. Elevated CO<sub>2</sub> concentrations were provided by the mixture of industrial high CO<sub>2</sub> and ambient CO<sub>2</sub>. 500 and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> were obtained by adjusting the velocity and amount of flow of industrial CO<sub>2</sub> and ambient CO<sub>2</sub>. CO<sub>2</sub> concentration was monitored by the CI-301 gas analyzer once a week. The plants were four years old, with average height of 47 cm, which are daily irrigated except rainy day.

All measurements were made on the current-year needle. The measured needles were near the top crown and received the full sunlight. The experiment was begun after the plants had been exposed to the CO<sub>2</sub> treatments for 3 months in the fourth growing season. Seedlings had been treated by high CO<sub>2</sub> concentrations (500 and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>) continuously (24 h·d<sup>-1</sup>) during growing season from June to September since 1999.

#### Measurement of stomatal conductance and $c_i/c_a$ ratio

Stomatal conductance ( $g_s$ ) and  $c_i/c_a$  ratio were measured with a portable photosynthetic analyzer equipped with a conifer cuvette (LI6400, Li-Cor, Inc., Lincoln, NE). Reciprocal transfer experiment of  $g_s$  and  $c_i/c_a$  in high- and low-[CO<sub>2</sub>]-grown plants was carried out at three levels of CO<sub>2</sub> concentrations (350, 500 and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$ ) in the cuvette, respectively. All measurements were made directly under light saturating conditions. The readings were taken after allowing  $g_s$  to reach a steady state.

#### Stomatal number

Twenty needles were collected at random from 20 plants per treatment. Stomatal number was separately counted on the upper and lower surface of current needles. Upper and lower surface of each needle were cut down 3-mm long epidermis along the needle, which was viewed with a microscope. The number of all stomatal lines and number of stomata per line on 3-mm long epidermis were counted.

#### Statistics

Mean values of stomatal conductance and  $c_i/c_a$  ratio were compared separately. One-way analysis of variance was performed for three comparisons. One contrast was carried out to compare needles grown and measured at 350  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> with those grown at 700 and 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> but measured at 350  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>. The second contrast was to compare needles grown and measured at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> with those grown at 700 and 350  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> but measured at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>. The third contrast was to compare needles grown and measured at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> with those grown at 500 and 350  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> but measured at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>. Stomatal line and stomatal number on upper, lower surface and whole needle were compared among the four treatments. All statistical tests were performed using SPSS 11.5 software. The conclusions were reached by the LSD tests.

## Results

#### Reciprocal transfer experiment of $g_s$

Stomatal conductance ( $g_s$ ) of *Pinus sylvestris* in the 500

$\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> was 61% and 4% higher than those in control chamber and on un-chambered field when measured at their respective growth CO<sub>2</sub> concentrations. Similarly,  $g_s$  in 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> increased by 11% and decreased by 28%, compared with those of growing in the control chamber and on un-chambered field, respectively (Table 1). The difference was significant between elevated CO<sub>2</sub> and ambient CO<sub>2</sub> ( $p < 0.05$ ).

When measured at 500 or 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>, stomatal conductance of *Pinus sylvestris* at elevated CO<sub>2</sub> concentrations was substantially higher than those at ambient CO<sub>2</sub> concentration.  $g_s$  at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> was 22% lower than that on un-chambered field when measured at 350  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>.  $g_s$  at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> was the highest at any measurement CO<sub>2</sub> concentration.  $g_s$  of both high-[CO<sub>2</sub>]-grown and control plants declined with the increase of measurement CO<sub>2</sub> concentration.

**Table 1.** Mean stomatal conductance ( $\text{mol} \cdot \text{m}^{-2} \cdot \text{s}^{-1}$ ) in *Pinus sylvestris* grown at ambient CO<sub>2</sub> and elevated CO<sub>2</sub> concentrations measured at three different CO<sub>2</sub> concentrations (350, 500, and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>)

Growth conditions ( $\mu\text{mol} \cdot \text{mol}^{-1}$ CO <sub>2</sub> )	Measurement CO <sub>2</sub> concentration ( $\mu\text{mol} \cdot \text{mol}^{-1}$ )		
	350	500	700
700	0.284±0.001	0.273±0.0002	0.262±0.0004
500	0.399±0.001	0.379±0.001	0.371±0.002
Control chamber (350)	0.235±0.001	0.216±0.0003	0.206±0.0003
Un-chambered field (350)	0.365±0.004	0.235±0.001	0.181±0.003

**Note:** Values shown above are means ± standard error. Comparisons were made among the four treatments at each measurement CO<sub>2</sub> concentration. Results for a one-way analysis of variance showed the difference were significant ( $p < 0.05$ ).

#### Reciprocal transfer experiment of $c_i/c_a$ ratio

When measured at their respective growth CO<sub>2</sub> concentration, high-[CO<sub>2</sub>]-grown plants exhibited lower  $c_i/c_a$  ratios compared with the control plants (Table 2).  $c_i/c_a$  ratio of *Pinus sylvestris* grown at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> was 5% and 3% lower than those at control chamber and un-chambered field, respectively. It was 4% and 2% lower than the control chamber and un-chambered field for *Pinus sylvestris* grown at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>. The difference was significant between elevated CO<sub>2</sub> and ambient CO<sub>2</sub>. However, the  $c_i/c_a$  ratio increased for *Pinus sylvestris* grown in high CO<sub>2</sub> concentrations when exposing to 350  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>.

**Table 2.**  $c_i/c_a$  ratio of *Pinus sylvestris* grown at ambient and elevated CO<sub>2</sub> concentrations measured at three different CO<sub>2</sub> concentrations (350, 500 and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>), respectively

Growth conditions ( $\mu\text{mol} \cdot \text{mol}^{-1}$ CO <sub>2</sub> )	Measurement CO <sub>2</sub> concentration ( $\mu\text{mol} \cdot \text{mol}^{-1}$ )		
	350	500	700
700	0.719±0.003	0.685±0.001	0.667±0.001
500	0.728±0.001	0.676±0.001	0.693±0.001
Control chamber (350)	0.701±0.002	0.635±0.002	0.600±0.001
Un-chambered field (350)	0.687±0.002	0.686±0.0003	0.660±0.002

**Note:** Values shown above are means ± standard error. Comparisons were made among the four treatments at each measurement CO<sub>2</sub> concentration. Results for a one-way analysis of variance showed the difference were significant except the comparison between 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> and un-chambered field at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> measurement CO<sub>2</sub> concentration ( $p < 0.05$ ).

When control plants grown at ambient CO<sub>2</sub> concentration were measured at high CO<sub>2</sub> concentrations (500 and 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>) the  $c_i/c_a$  value decreased. The  $c_i/c_a$  ratios of high-[CO<sub>2</sub>]-grown plants were higher than those of control plants when measured at the same CO<sub>2</sub> concentration.

### Stomatal number

Stomata of *Pinus sylvestris* occur in a few of straight lines running along the length of the needle on both sides of the needle. The number of stomatal line per needle (including upper and lower surface) at high CO<sub>2</sub> concentrations was significantly lower than that on un-chambered field (Table 3). The number of stomatal line at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> approximately equals to that at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>. The number of stomatal line of plants grown on un-chambered field was higher than that in the control chamber though both accepted ambient CO<sub>2</sub> concentration. The stomatal line of plants in the control chamber was 10% higher than those at 700 and 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>. But there was no significant differences between control chamber and elevated CO<sub>2</sub> concentrations. The allocation of stomatal line and number of stomata was different between upper and lower surface of needle for high- and low-[CO<sub>2</sub>]-grown plants. Stomatal line and number of stomata on upper surface were more than those on

lower surface. The number of stomatal line on the upper surface of needle grown at 700 and 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> decreased by 16% and 8%, respectively, compared with that in the control chamber. Similarly, the number of stomatal line on the upper surface of needle grown at 700 and 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> decreased by 25% and 19%, respectively, compared with that on un-chambered field. The number of stomatal line on the lower surface of needle grown at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> showed no reduction and was 11% lower grown at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>, compared with that in the control chamber. *Pinus sylvestris* grown at 700 and 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> exhibited that the numbers of stomatal line on the lower surface of needle were 11% and 23% separately lower than that at the un-chambered field. There was no significant difference on the number of stomata on lower surface among four treatments. Stomatal number per unit long needle on upper surface at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> was much higher (increased by 16%) than that on the un-chambered field. However, elevated CO<sub>2</sub> did not significantly change the total stomatal number (including upper and lower surface). The results of variance on the stomatal line and stomatal number among four treatments were shown at Table 4.

**Table 3. Stomatal line and stomatal number of current-year needle of *Pinus sylvestris* exposure to high CO<sub>2</sub> concentrations for four growing seasons**

Indexes	Growth CO <sub>2</sub> concentration ( $\mu\text{mol} \cdot \text{mol}^{-1}$ CO <sub>2</sub> )			
	700	500	Control chamber (350)	Unchambered field (350)
Number of stomatal line on upper surface of needle	7.6±0.413	8.3±0.442	9.0±0.397	10.2±0.485
Number of stomata 1mm long needle on upper surface	12.1±0.252	11.4±0.249	11.7±0.246	11.3±0.163
Number of stomatal line on lower surface of needle	6.3±0.448	5.5±0.256	6.2±0.296	7.1±0.352
Number of stomata 1mm long needle on lower surface	10.8±0.171	10.7±0.229	11.0±0.256	10.8±0.183
Number of stomatal line per needle	13.9±0.737	13.8±0.627	15.2±0.601	17.3±0.781
Number of stomata 1mm long needle	22.9±0.359	22.1±0.389	22.6±0.443	22.2±0.294

**Table 4. Results of one-way analysis of variance of stomatal line and stomatal number (P: 0.05 level)**

	1-2	1-3	1-4	2-3	2-4	3-4
Number of stomatal line on upper surface of needle	0.227	0.021	0	0.259	0.003	0.055
Number of stomata 1mm long needle on upper surface	0.056	0.231	0.034	0.464	0.832	0.345
Number of stomatal line on lower surface of needle	0.106	0.838	0.129	0.156	0.002	0.086
Number of stomata 1mm long needle on lower surface	0.693	0.537	0.923	0.315	0.766	0.476
Number of stomatal line per needle	0.959	0.171	0.001	0.156	0.001	0.039
Number of stomata 1mm long needle	0.156	0.591	0.167	0.385	0.971	0.405

Note: 1: 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>; 2: 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub>; 3: control chamber; 4: un-chambered field

### Discussion

Stomatal conductance, ratio of intercellular to ambient CO<sub>2</sub> concentration and stomatal number are main parameters of assessing stomatal behavior at elevated CO<sub>2</sub> concentration. Stomatal conductance can well describe the dynamic changing trend of stomatal characteristics, while relative stable properties can be provided by stomatal number (Zhang *et al.* 2002).

It is widely stated that elevated CO<sub>2</sub> concentration will cause the reduction of stomatal conductance. Stomatal conductance is affected primarily by stomatal aperture and the number of stomata i.e. stomatal density (Weyers and Lawson 1997). Thus changes in size and number of stomatal aperture play key roles in stomatal conductance. Since stomatal number of *Pinus sylvestris*

was decreased by elevated CO<sub>2</sub> concentrations, the increase of stomatal conductance at elevated CO<sub>2</sub> concentrations mainly related to stomatal aperture. *Pinus sylvestris* grown at 500  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> showed the highest stomatal conductance at any measuring CO<sub>2</sub> concentration. However, the change of stomatal conductance of *Pinus sylvestris* grown at 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> was related to the measuring CO<sub>2</sub> concentration. Stomata are sensitive to some environmental stimuli, particular light, humidity and CO<sub>2</sub>. Therefore, changes of stomatal conductance in 700  $\mu\text{mol} \cdot \text{mol}^{-1}$  CO<sub>2</sub> could be caused mainly by the change of environmental CO<sub>2</sub> concentration. Therefore, the simplest interpretation is that the difference in stomatal conductance between high- and ambient-[CO<sub>2</sub>]-grown plants was a result of the direct adjustment of stomatal aperture.

The change of stomatal aperture or conductance could affect

the  $c_i/c_a$  ratio of stomata which are directly sensitive to intercellular  $\text{CO}_2$  concentration (Mott, 1990).  $g_s$  and  $c_i/c_a$  ratio in high- $[\text{CO}_2]$ -grown plants were higher than those of control plants when measured at high  $\text{CO}_2$  concentrations, confirming that stomata of *Pinus sylvestris* acclimated to long-term exposure to high  $\text{CO}_2$  concentrations. High- $[\text{CO}_2]$ -grown plants showed a decrease for the ratio of  $c_i/c_a$  compared with control plants when measured at their respective  $\text{CO}_2$  concentration of growth. The decrease of  $c_i/c_a$  ratio in high- $\text{CO}_2$ -grown plants was mainly caused by higher ambient  $\text{CO}_2$  concentration (700 and  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$ ).

Given no adjustment to the change of increasing atmospheric  $\text{CO}_2$  concentration the  $c_i/c_a$  ratio would remain constant. However, Ellsworth (1999) found a similar tendency in  $c_i/c_a$  ratio for well-watered *P. taeda*. Sage (1994) and Drake *et al.* (1997) also demonstrated it in many experiments, including those in which plants were grown for long periods in high  $\text{CO}_2$  concentration. But it is surprising that the ratio of  $c_i/c_a$  unchanged at high  $\text{CO}_2$  concentration. Sage (1994) found that except under water and humidity stress,  $c_i/c_a$  exhibited inconsistent change in a variety of  $\text{C}_3$  species. In our study, high- $[\text{CO}_2]$ -grown *Pinus sylvestris* exhibited that  $c_i/c_a$  ratio was below the control plants when measured at their respective growth condition, as also demonstrated by Wong (1993).  $c_i/c_a$  ratio increased in high- $[\text{CO}_2]$ -grown plants when measured under 700 and  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$ , confirming that stomata do not always maintain  $c_i/c_a$  constant.

Stomatal number of current-year needle of *Pinus sylvestris* decreased under 700 and  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$ , and the allocation pattern of stomata between upper and lower surface of needle gave rise to change. Stomatal lines on the upper surface of needle showed larger reduction at  $700 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$ , and that on the lower surface showed larger decline at  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$ . The difference was not significant though the total stomatal lines of whole needle (including upper and lower surface) in the control chamber were higher than that in elevated  $\text{CO}_2$ . The total stomatal lines of needle on un-chambered field were remarkably higher than those in the open top chambers (including 700,  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$  and control chamber). Therefore, the microenvironment of open top chamber can affect the number of stomatal line of *P. sylvestris*. That is both elevated  $\text{CO}_2$  and microenvironment of open top chamber decreased the number of stomatal line, but they did not change the stomatal number per unit long needle. Therefore, the total stomatal number of *P. sylvestris* had a decrease at 700 and  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$  by decreasing stomatal line on upper and lower surface of needle. The change of stomatal number is a result of long-term exposure to high  $\text{CO}_2$  concentrations. The decline of stomatal number for *Pinus sylvestris* did not significantly affect the change of stomatal conductance. Some studies showed stomata density did not change at high  $\text{CO}_2$  concentration.

There was difference in stomatal response for *Pinus sylvestris* to 700 and  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$ . The increases of stomatal conductance and  $c_i/c_a$  ratio of plants grown at  $500 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$  were relatively bigger than those at  $700 \mu\text{mol} \cdot \text{mol}^{-1} \text{CO}_2$ . In addition, the allocation of stomatal line on upper and lower surface was different, too. Stomatal behavior of plants in the control chamber was also different from that on

un-chambered field. By this study, we found that the microenvironment of open-top chamber affected the physiological characteristics of *Pinus sylvestris*. But we still do not confirm which factors and how these factors operate.

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